## EFFECTS OF MARINATING PROCESS ON MUSSELS PHYSICOCHEMICAL AND MICROBIOLOGICAL QUALITY ATTRIBUTES DURING REFRIGERATED STORAGE

### Héla CHERIFI et Saloua SADOK

Laboratoire de Biotechnologie Bleu et Bioproduits Aquatiques (B3Aqua), INSTM , centre La Goulette, Port de Pêche 2060, Tunis, TUNISIE, Email/ \* hela.cherifi@hotmail.fr; salwa.sadok@instm.rnrt.tn

#### RESUME

En Tunisie, la moule *Mytilus galloprovincialis* est un bivalve peu consommé par les tunisiens à cause du manque de conscience de sa valeur nutritive. Le but de cette étude est d'améliorer l'image de ce produit sur le marché en utilisant une nouvelle méthode de conservation à savoir le marinage. Durant cette étude, le procédé de marinage utilisant l'acide acétique, le sel et l'huile végétale a été adopté afin de prolonger la durée de vie du produit tout en conservant sa qualité nutritionnelle. L'effet synergique des différents composants de marinade (acide acétique, le sel et l'huile végétale) a été évalué sur la qualité physico-chimiques (pH et l'activité de l'eau), microbiologique, chimique, (ABVT, TBA) et biochimique des moules marinées emballées sous vide et conservées 15 jours dans la glace à 4 ° C. Les résultats obtenus pour les moules marinées ont été comparés avec les résultats d'un lot contrôle de moules. Les teneurs en Azote Basique Volatil totale (ABVT) et en acide thiobarbiturique (TBA) ont été en dessous des valeurs limites seuils d'acceptabilités (25mg / 100g et 3 mg MDA / kg) au cours de la période de stockage. Les teneurs en ABVT et en TBA étaient de 14,3  $\pm$  0,006 mg/ 100g et 0,2 mg MDA / kg respectivement pour le lot de contrôle et 14,12  $\pm$  0,02 mg / 100g et 0,3 mg MDA / Kg pour lot mariné. Le nombre de la flore mésophyle totale des moules marinées (3,21 log UFC / g) était significativement plus faible par rapport au moules témoins (4,14 log UFC / g). Les analyses effectuées ont déterminé une DLC de 15 jours pour les deux lots avec une détérioration de la qualité nutritionnelle nettement plus lente pour le lot mariné.

*Mots clés:* Moules marinées, *Mytilus galloprovincialis*, Composition biochimique, Qualité microbiologique, Qualité physico-chimique

#### ABSTRACT

In Tunisia, mussels *Mytilus galloprovincialis* are not familiar seafood because of cultural influence and convenience aspects. The aim of the present study is to develop a ready to eat seafood product as a novel variety of mussel. Marinating process using acid, salt and vegetable oil was chosen in order to achieve a longer shelf life, to improve taste and to maintain nutritional quality of the product. Synergistic effect of different pickling agents (acetic acid, salt and vegetable oil) was evaluated on physicochemical (pH and water activity), microbiological, chemical (TVB-N and TBA) and biochemical (water, ash, proteins, lipids and carbohydrates) quality of marinated mussels *Mytilus galloprovincialis* during vacuum-packaged storage during 15 days at 4°C. Obtained results of marinated mussels samples were compared with results of a control mussels samples. Total Volatile Basic Nitrogen (TVB-N) and Thiobarbituric acid (TBA) values were determined below the thresholds of acceptability limits values (25mg/ 100g and 3 mg MDA /kg respectively) during the storage period. TVB-N and TBA values were found as 14.3  $\pm$  0.006 mg / 100g and 0.2 mg MDA/kg respectively for control mussel and 14.12  $\pm$  0.02 mg / 100g and 0.3 mg MDA / Kg for marinated mussels. (3.21 log CFU / g) had significantly lower level compared to control group (4.14 log CFU/ g). Analysis evaluated a shelf life of 15 days for both batches with a significantly slower deterioration of the quality for the marinated batch.

Key words: Marinated Mussels, Mytilus galloprovincialis, Biochemical content, microbiological quality, physicochemical quality

### **INTRODUCTION**

In Tunisia, shellfish farming is mostly limited to the lagoon of Bizerte, located in the North of the country. Presently, this production is valued at an average of 1635 tons (DGPA, 2015). Because of several aliases such as the increase of water temperature in summer, significant mortality of livestock is observed (ie, current year) especially for mussels *Mytilus galloprovincialis*, causing great economic losses for producers. Furthermore, in some periods of the year,

alerts of biotoxins prohibit harvesting, marketing and consumption of the product. On the other hand, when mussels are normally produced, they are exclusively sold fresh for limited market such as superstores and restaurants (DGPA, 2015). As an alternative of a breakthrough, could be processing unsold mussels during high production turning it into convenient seafood with various applications and many new marketing opportunities. The development and application of innovative and efficient processing and preservation techniques are essential to produce high added and nutritional value products (Aveiro et al., 2007).

Such practice would not only ensure marketing and large distribution of the product during the periods of high and low mussel's production, but also encourage its wider aquaculture off the lagoon, in the open sea for a regular market supply. Mussels have high nutritional values making them ideal nutrients in human diet. Consumption of this bivalve helps to provide polyunsaturated fatty acids with known health beneficial effects, including essential vitamins, proteins with high biological value as well as minerals (Orban et al., 2002). This product is also known as a shellfish product with low fat and cholesterols contents (Erkan, 2005). Processed products, however must, fulfil the requirements of consumers, to be attracting, easy to prepare and good to be tasted.

Taking into account all cited considerations, and within SecurAqua project; the INSTM research group focused efforts to elaborate added-value product using mussels from the lagoon of Bizerte.

Numerous studies (Dalgiç & Erkoyuncu 2003; Kyriazi-Papadopoulou et al., 2003; Şengör et al., 2004; Goulas et Kontominas, 2005,; Ozgul & Balikci, 2013; Maktabi et al., 2015) have been done on various processing methods in order to preserve the nutritional quality of the seafood products while improving their shelf life.

Marinating is among technique used in food preserving. This technique is based on the treatment of the product with various solutions using salt, spices, lemon's juice, and numerous other compounds as pickling agents in order to ameliorate the organoleptic properties and tenderize the texture and structure of the product (Yashoda et al., 2005). Moreover, marinating reduces the bacterial and enzymatic activity, contributes to the improvement of sensory qualities of the product and ensures its extended but limited shelf life (Sallam et al., 2007).

Generally, the marinated seafood are treated with acids (acetic acid), sugar, salt, spices and oil in order to improve tenderness, juiciness and the flavour of the product's flesh (Hwang & Tamplin, 2005). Several studies were interested in the evaluation of the effect of the marinades on fishes' nutritional and microbiological quality (Goulas et Kontominas, 2005; Kilinc & Cakli, 2005; Cadun et al., 2008), however limited studies were carried out concerning marinated bivalves especially mussels.

In this work an interest was given to the evaluation of the effect of a mixture of salt, acetic acid and vegetable oil on the microbiological, physicochemical, and biochemical quality characteristics of the marinated mussels *Mytilus galloprovincialis* during vacuum-packaged storage during 15 days at  $4^{\circ}$ C

## MATERIALS AND METHODS

#### Collection and preparation of the samples

Fresh mussels weighing about 8 kg were provided from shellfish company located in the lagoon of Bizerte, North Tunisia. On arrival to the laboratory, mussels were well cleaned, and byssi's thread was removed. All specimens were kept at 4°C until further processing.

#### Marinating process

Following shell cleaning, depurated mussels were cooked until all shells were open (5 to 10 minutes of cooking). Mussels which remained closed were eliminated (Turan et al., 2006). After cooking, marinating process were performed. The different steps of the marinating process are presented in table I

Steps	Marinating process				
Step 1	Cooking and collecting mussel's meat				
	Dividing mussels in 2 batches				
Step 2	Batch 1: control mussels	Batch 2: marinated mussels			
Step 3	No application of the marinating process.	<ul> <li>Application of the marinating process:</li> <li>Adding salt</li> <li>Rinsing with vinegar</li> <li>Soaking in vegetable oil</li> </ul>			
Step 4	Sterilization: at 121°C for 20 min				
Step 5	Storage under vacuum packaging and conservation for 15 days in ice at 4°C				

 Table I: Marinating process applied to the mussel Mytilus galloprovincialis

During the storage period, samples were taken for analyses every three days (day 0, 3, 6, 9, 12 and 15) from control and marinated batch. For each sample, three bags of vacuum-packed mussels (n=30g/ bag) were taken and samples were well homogenised and preserved at -80°C for chemical analyses. Microbiological analyses, water activity and pH determination were performed the same day of the sampling.

#### Physical and chemical analysis Water activity pH and measurement

During this study, pH value of mussel's meat was measured using pH indicator paper CARLO ERBA. The water activity (Aw) was measured using the LabSwift-aw system novasina (The art of Precision Measurement).

#### Microbiological analysis

Microbiological quality was evaluated according to the method of Harrigan and McCance (1976). For each sample, 10g were taken and homogenized in 90 ml peptone water. Decimals dilutions were prepared from  $10^{-1}$  dilution. Total viable count was determined using Plate Count Agar as the medium. Plates were incubated at 26°C for 24-48 h. Analysis was made in triplicate in each sample.

### **Chemical analysis**

The total Volatile Basic Nitrogen (TVB-N) content was determined according to the method of Ruiz-Capillas & Horner (1999) using the system of « Flow injection analysis » (FIA). Thiobarbituric acid (TBA) was evaluated using the method of Genot (1996). TBA values were expressed in units of mg/malonaldehydre /Kg sample.

#### **Proximate composition**

Evaluation of water and ash content was performed according to the method of AOAC (1990) Water content was determined at 105°C until a constant weight was obtained. Ashes were evaluated by ignition of the dry sample in an oven at 550°C during 6 hours. Total proteins content was determined according to the method of Lowry modified by Hartree (1972) and lipid content was determined by the method of Folch *et al.* (1957). All of these methods were submitted to harmonisation with the procedures used in CUPT (Consortium Universitaire de Trapani) laboratory within BIOVecQ project.

#### Statistical analysis

All obtained results were reported as Mean  $\pm$  SD. One way ANOVA test followed by the least significant difference test (LSD) in the statistical software program SPSS were used to evaluate any significant difference (p< 0.05). The software used was SPSS version 17.0.

## **RESULTS AND DISCUSSION**

# Evaluation of pH and water activity in control and marinated mussels samples during the storage period at $4^{\circ}C$

High pH level in food is one of the main spoilage indicators in marinated products. The increase of pH levels, because of the nitrogenous compounds production by bacteria, indicates the loss of quality of the product and its spoilage (ICMSF, 1993).

The evolution of pH values in the control and marinated mussels samples during the refrigerated storage period is summarized in Figure 1. Initial pH value obtained for cooked mussels was 7.5; this result is compared to data reported by Turan *et al.* (2008) for boiled mussels.

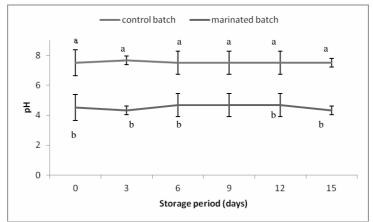
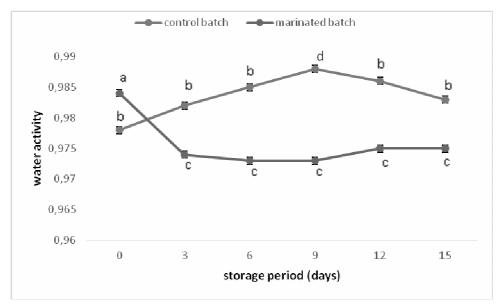


Figure 1: Evolution of pH value in control and marinated mussels samples stored under vacuum packaging during 15 days at (4°C). Bar = standard error; n = 6 in each case; different letters= mean values for each group are significantly different (p<0.05).

However, marinated mussels' flesh showed a significant lower pH values compared to those obtained for the control samples. This difference is due to the addition of the acetic acid during the marinating process (Gopal et al., 1985). Similar results were reported by Sengör et al (2004), Maktabi et al (2015), and Ozgul et al (2010) for smoked marinated mussels, marinated rainbow trout fillet and smoked marinated anchovy respectively. Marinating process performed to cooked mussels using acetic acid and salt aimed to ensure the stability and the extension of shelf life of the product. According to several studies, acetic acid has an effective antimicrobial activity (Aveiro et al., 2007). Throughout the 15 days of refrigerated storage, the pH values in control and marinated mussels' samples remained unchanged.

According to McLay (1972), food poisoning and growth of spoilage bacteria may occur when pH values exceed 4.8. In the present study, pH values in marinated mussels remained far below 4.6 throughout the period of storage. However in control mussels, pH values were > 7, which may favour the development of bacteria.

The physical, chemical and microbiological stability of food is strongly depends on the water content of the product. Therefore the concept of water activity (aw) was adopted as a reliable evaluation method of the microbial growth, non enzymatic and enzymatic activities and lipids oxidation (Rahman & Labuza, 1999). In this study, results related to the evolution of water activity in control and marinated mussels samples during the storage period at 4°C are illustrated in Figure 2.



**Figure 2**: Evolution of water activity content in control and marinated mussels samples stored under vacuum packaging during 15 days at (4°C). Bar = standard error; n= 6 in each case; different letters= mean values for each group are significantly different (p<0.05).

According to conventional values (Ballesteros et al., 1993), both mussels' batches in this study, can be considered to be food products with high water activity limits as the recorded aw values were  $\geq 0.86$ . However, water activity in marinated mussels showed a significant drop after 3 days of storage to remain significantly lower than values found in the control mussels' batch (p < 0.05) for the rest of the storage. Such aw decrease can be attributed to salt effect, acting as depressor agent of water activity during the maturation phase (Girard, 1988). Indeed, the water activity decreases as the sodium chloride concentration increases (Sainclivier, 1985). It is assumed that the measured values of water activity generally correlate well with metabolic activity and potential growth of microorganisms (Gould, 1985;

Gould et Christian 1988). According to several studies, each microorganism has a critical water activity below which growth cannot occur (Leistner et al., 1981; Beuchat, 1983). For example pathogenic bacteria have a critical water activity varying from 0.85 to 0.86 (Silverman et al., 1983; Ballesteros et al., 1993). The obtained results showed that the addition of salt in marinating process ensure a significant lower water activity in marinated mussels' flesh compared to control batch and as a result a better conservation of the product from microorganisms activity. However, further investigation on salt/flesh ratio during marinating process is needed to assure lower water activities for better effects on the product quality with longer shelf life.

# Evaluation of the microbiological quality in control and marinated mussels samples during the storage period at $4^\circ C$

Microorganism's proliferation is among the most important factors in the deterioration of food with special emphasis to seafood products as they are more alterable (Maktabi et al., 2015). Figure 3 presents the enumeration of the total mesophilic flora in control and marinated mussels samples during refrigerated storage.

Initial number of total mesophilic bacteria was 3.04 log CFU/G and 2.96 log CFU/g for control and marinated samples respectively. These results are in agreements with those found for other marinated seafood products (Stamatis & Vafidis, 2009; Ozgul & Balikci, 2013).

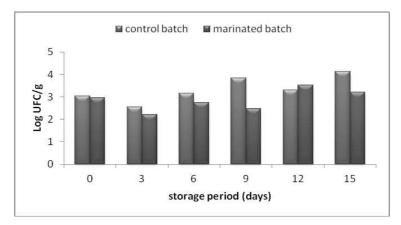
In the present study, the initial microbial loads for control and marinated mussels samples were < the limits  $(10^6 \text{ CFU/g})$  established by International Regulation (ICMSF, 1986) and adopted by the DGSV (2011). Such low initial counts indicate the good quality of the raw product. During storage, an increase among total mesophilic bacteria was observed to reach 4.14 log CFU/g for the control samples and 3.21 log CFU/G for the marinated samples.

Compared to several other studies, both mussels' batches were qualified as "good" food product since

they did not exceed the upper acceptability limit (Huss, 1988; Bilgin et al., 2006; Cakli et al., 2006; Bao et al., 2007; Duyar et al., 2012). Moreover, marinated mussels' batch presented lower total mesophilic flora than that of the control batch during the 15 days of storage. These results are in agreements with those found in the literature (Maktabi et al., 2015; Kilinc & Cakli, 2004). This result could be explained by the synergic effect of salt and vinegar added during marinating process, which inhibited the proliferation of micro-organisms (Jarvis et al., 1987). However, the increasing of total mesophilic bacteria during storage for marinated batch suggest that the acetic acid/salt ratio added during marinating process was not enough to protect the product, and further investigation on the adequate ratio should be done.

# Evaluation of the chemical indices (TVB-N and TBA) in control and marinated mussels samples during the storage period at 4°C

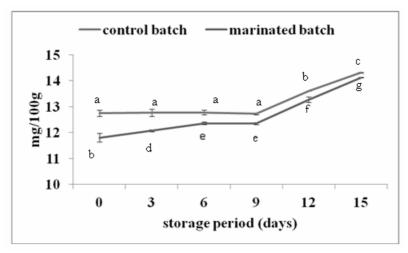
The evaluation of Total Volatile Basic Nitrogen (TVB-N) and Thiobarbituric acid (TBA) are used for the determination of the spoilage level of seafood during storage period. As freshness indices, monitoring of TVB-N and TBA was carried out during 15 days of refrigerated storage.



**Figure 3:** Enumeration of the total mesophilic flora (26°C) in control and marinated mussels samples stored under vacuum packaging during 15 days at 4°C (n= 3 in each case)

# Evaluation of the Total Volatile Basic Nitrogen (TVB-N) content

The evaluation of TVB-N content is one of the most used methods of routine seafood quality determination. High TVB-N content is an indicator of deterioration of the product caused by both bacterial activities and endogenous enzymes (Ruiz-Capillas & Moral, 2005). In this study, the variation of TVB-N levels in control and marinated mussels samples under refrigerated storage are represented in figure 4.



**Figure 4:** Evaluation of Total Volatil Basic Nitrogen (TVB-N) (mg/100g) content in vacuum packed control and marinated mussels samples stored during 15 days at 4°C. Vertical Bar= Standard Error, (n= 6 in each case); Different Letters= mean values for each group are significantly different (p<0.05).

Initial values of the total volatile basic nitrogen content in control and marinated samples were  $2.75 \pm 0,001$  and  $1.80 \pm 0,016$  mg/100g respectively. These results are in agreement with those mentioned by Turan et al., (2006).

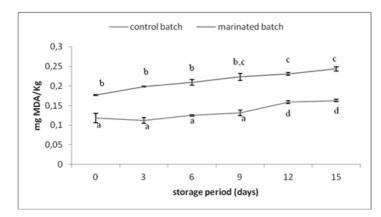
A significant increase was observed for the two batches from the 9<sup>th</sup> day of storage with significantly lower values for marinated mussels' samples compared to control samples. Obtained results are in agreement with those mentioned in the literature (Kilinc & Cakli, 2004; Ozyurt et al., 2012). Koutsoumanis & Nychas, 1999 and Ruiz-Capillas & Moral (2001) explain this increase by spoilage bacteria activity. Huss (1988) suggested different categories for seafood products quality according to its TVB-N content: samples with a content of 25 mg/ 100 g of TVB-N are considered as "very good", for a content of 30 mg/100 g of TVB-N product is regarded as "good", for a content of 35 mg/100g product is defined as "negotiable", and if the content is higher than 35 mg / 100 g TVB-N product is considered as "rotted".

During this study, reached values in the 15<sup>th</sup> day of storage were as followed:  $4.3 \pm 0,012 \text{ mg}/100 \text{ g}$  for the control samples and  $4.12 \pm 0,006 \text{ mg}/100\text{ g}$  for the marinated samples, however content of TVB-N in both batches remained largely lower than the higher

limits of acceptability 25 mg/100g (CEC, 1995). Erkan (2005) and Goulas et Kontominas (2005) suggested values limits of acceptability for mussels of about 15 and 22-25 mg/100g respectively. Results of TVB-N content for both batches obtained during this work remained lower than the limits of acceptability mentioned by last cited works.

# Evaluation of thiobarbituric acid (mg MDA/Kg) content

The content of thiobarbituric acid in seafood reflects the degree of lipid degradation corresponding to products of the secondary step of lipids oxidation. The oxidation of lipids involves considerable changes in the organoleptic properties of seafood as in the texture and the flavor (Aubourg et al., 2005, Brannan et Erickson, 1996; Pirini et al., 2000, Regost et al., 2004). This oxidation, is dependent on the presence of oxygen in the environment of storage, of the temperature of storage, the presence of pro-oxidants like the metal ions, and of the lack of natural antioxidants (like a-tocopherol) or of its deterioration in the flesh of the product (Huss, 1995). As a good indicator of food quality (Tarladgis et al., 1960; Vareltzis et al., 1993), the variation of the content of TBA recorded for control and marinated batches is represented on figure 5. Initially TBA content results obtained for control and marinated mussels samples were about 0.12  $\pm$  0.01 mg MDA /Kg for control mussels and 0.17mg MDA/kg for the marinated mussels. Ozgul & Balikci (2013) and Cadun et al. (2005) also reported low contents of TBA for marinated fishes and shrimps respectively. During this study, a significant increase of the contents of TBA was observed from the 12<sup>th</sup> day of storage for control and marinated samples to reach 2 and 3 mg MDA/kg respectively. Similar results of increased TBA values during storage are mentioned by numerous studies done on the effect of marinating process on seafood during refrigerated storage (Ozgul et al., 2010; Maktabi et al., 2015). However, obtained results in the present work show significant higher lipid oxidation in marinated mussels' samples compared to control mussels. These results can be



**Figure 5:** Evaluation of Thiobarbituric acid (TBA) (mg MDA/Kg) content in vacuum packed control and marinated mussels samples stored during 15 days at 4°C. Vertical Bar= Standard Error, (n= 6 in each case); Different Letters= mean values for each group are significantly different (p<0.05).

explained by higher rates of lipids in marinated mussels due to the addition of vegetable oil during marinating process and in consequence a greater susceptibility to their oxidation. Similar results were reported in other studies (Pakawatchai et al., 2009; Pezeshk et al., 2012, Maktabi et al., 2015) for marinated seafood products. According to several studies, the rate of TBA which is used for the determination of the rancidity of oil must be lower than 3 mg MDA/kg in a food defined as a very good quality product and should not exceed 5 mg MDA /Kg in food of good quality (Varlik et al., 1993). Other studies reported that rancidity starts in food products when TBA values exceeds 4 Mg MDA/Kg and that the limit beyond which the seafood products will develop an odour and/or unpleasant taste and become unsuitable for consumption is in the order of 7-8 mg MDA/Kg (Kaya & Basturk., 2015). Data in the present study suggest that the contents of TBA in both mussel's batches remained lower than the threshold of acceptability mentioned by literature (Kaya & Basturk., 2015).

Biochemical content in control and marinated mussels samples during the storage period at  $4^{\circ}C$ 

Biochemical composition of bivalves is strongly related to water temperature, food availability and the gametogenic cycle of animals (Small & van Stralen, 1990). Processors have a direct interest in the biochemical composition of product, needing to know the nature of the raw material before different manufacturing techniques can be correctly applied (Murray & Burt, 1969). Monitoring of biochemical composition of raw material and marinated mussels was carried out during 15 days of refrigerated storage.

# Characterization of the initial proximate composition

The quality and state of freshness of a finished product depend directly on the initial quality of the raw product. A characterization of the initial proximate composition of samples from the two batches of mussels was carried in order to evaluate occurring nutritional changes during vacuumpackaged storage during 15 days at 4°C (table II).

Tableau II: Com	parison of initial	proximate com	position of control	and marinated	samples

Proximate composition	Control batch (day 0)	Marinated batch (day0)		
Ash content (g/100g)	$2.17\pm0.2^{a}$	$2.96 \pm 0.047^{b}$		
Moisture content (g/100g)	$80.38 \pm 0.09^{a}$	$77.21 \pm 0.21^{b}$		
Total protein content (g/100g)	$7.18 \pm 0.12^{ m a}$	$7.43 \pm 0.42^{\mathrm{a}}$		
Total lipid content (g/100g)	$2.40\pm0.05^{a}$	$7.58 \pm 0.06^{b}$		

Mean  $\pm$  SD; different letters= mean values for each group are significantly different (p<0.05); n=6 in each case

Results in Table II show that the ash and total lipids content were significantly higher (p < 0.05) in marinated mussels compared to control mussels.

Turan et al (2006) showed that ash content increased in cooked salted mussels due to the effect of salt addition. These results are explained by a loss of water from the flesh replaced in the marinated mussel's flesh by a salt penetration. The high lipid content in marinated mussels is explained by the addition of vegetable oil during the marinating process. Thus, this product is not recommended for people suffering from high cholesterol level in their blood.

# Monitoring biochemical changes during the storage period at $4^\circ C$

The evaluation of the proximate composition: water content, total ash content, total protein content, total lipid content and total carbohydrates content in control and marinated mussels samples was carried out during storage period at 4°C. The results are represented in table III.

Literature data on biochemical composition of the marinated mussels are very limited (Aveiro et al., 2007; Turan et al., 2006; Guldas & Hecer, 2010). The majority of elaborated studies were related to fish marinating process (Ozden, 2005; Kilinc & Cakli, 2004; Cabrer et al., 2002; Ozgul & Balikci, 2013).

During storage, the water and ash contents in control and marinated samples did not present significant variation. Similar results were found by Turan et al., (2006) which studied the effect of various methods of salting on the Mediterranean quality of the mussels *Mytilus galloprovincialis*.

**Table III:** Evaluation of the proximate composition changes in control and marinated mussels samples duringthe storage period at 4°C. Mean  $\pm$  SD; different letters= mean values for each group are significantly different(p<0.05); n=6 in each case</td>

Storage day		0	3	6	9	12	15
Analysis							
moisture	Control	$80.38^{a} \pm$	$79,2^{a} \pm 0.16$	$79.18^{\mathrm{a}}\pm0.09$	79.31 <sup>a</sup> ±	$80.41^{a} \pm$	$81.13^{a} \pm$
(g/100g)	batch	0,1			0.17	0.01	0.23
	Marinated	76.13 <sup>b</sup> ±	$77,97^{b} \pm$	$78.17^{b} \pm 0.12$	$78.33^{b} \pm$	$78.68^{b} \pm$	$78.53^{b} \pm$
	batch	0.15	0.27		0.03	0.22	0.12
Ash content	Control	$1.82^{a} \pm$	$1,85^{a} \pm 0.01$	$1.86^{a} \pm 0.03$	$1.84^{a} \pm$	$1.57^{a} \pm$	$1.11^{a} \pm$
(g/100g)	batch	0.15			0.002	0.06	0.08
	Marinated	$2.68^{b} \pm$	$2,12^{b} \pm 0.06$	$2.15^{b} \pm 0.083$	$2.67^{b} \pm$	$2.78^{b} \pm$	$1,121^{b} \pm$
	batch	0.15			0.02	0.12	0.04
Total protein	Control	$7.19^{a} \pm$	$3,6^{b} \pm 0.10$	$3.32^b\pm0.02$	$2.2^{d} \pm 0.06$	$2.53^{d} \pm$	$2.2^{d} \pm$
content g/100g	batch	0.12				0.04	0.04
	Marinated	$7.43^{a} \pm$	$4.95^{\rm c}\pm0.22$	$4.9^{c} \pm 0.09$	$3.2^{b} \pm 0.2$	2.84 <sup>b</sup> ±	3.06 <sup>b</sup> ±
	batch	0.04				0.06	0.003
Total lipid	Control	$2,4^{a} \pm 0.05$	$3^{b} \pm 0.07$	$3.86^{\circ} \pm 0.4$	$2.9^{b} \pm 0.08$	$2.96^{b} \pm$	$2.5^{a} \pm$
content	batch					0.015	0.03
(g/100g)	Marinated	$7,58^{d} \pm$	$4,82^{e} \pm 0.16$	$4.9^{e} \pm 0.10$	4.29 <sup>e</sup> ±	3.35 <sup>b</sup> ±	$4.66^{e} \pm$
	batch	0.06			0.11	0.05	0.25
Total	Control	$2,73^{a} \pm$	$2,35^{a} \pm 0.09$	$2.25^{a}\pm0.03$	$2.52^{a} \pm$	$2.52^{a} \pm 0.01$	2.01 <sup>a</sup> ±
carbohydrate	batch	0.06			0.04		0.007
content	Marinated	2,58 <sup>b</sup>	$2.52^{b} \pm 0.03$	$2.65^{b} \pm 0.03$	$2.27^{b} \pm$	2.21 <sup>b</sup> ±	$2^{b} \pm 0.03$
(g/100g)	batch	±0.07			0.02	0.09	

The contents of proteins presented a significant reduction for both batches of mussels during the first 3 days of storage then remained constant throughout the period of conservation. These results are comparable with those of Yeannes & Caslaes (2008) and Feeney (1977).

Lipid levels in marinated batches were significantly higher than levels observed in control batch. This difference can be explained by the addition of vegetable oil during marinating process.

The contents of carbohydrates did not show significant variation throughout the period of storage for both batches. Stamatis & Vafidis (2009) mentioned comparable results in marinated vacuum-packaged sea urchins during storage.

## CONCLUSION

The present study showed that:

- Compared to control mussels (cooked mussels); marinating using vinegar and salt induces low pH and water activity, reducing thus, the quality degradation of vacum-packed mussels stored at 4°C during 15 days.

- Marinated mussels have a mesophilic flora below the upper limits of acceptability threshold ( $10^6$ UFC / g) during 15 days of storage

- Marinated mussels present quality indices; TVB-N and TBARs were below the upper limits of acceptability of respectively 25 mg / 100 mg and 1-2 MDA / kg for 15 days of storage.

On the basis of all these analyses, a shelf life of 15 days was determined for the conservation of marinated mussels under vacuum packaging at 4°C.

This work contribute to the development of a range of value added seafood products which may be available in markets, making mussel more popular locally and may thus, play a great role in this endeavour. Beside the development of the international seafood value chain may created new market opportunities for mussel that stimulated local economy.

#### **AKNOWLEDGEMENTS**

This work was conducted as part of the IEVP- Cross border project **SecurAqua** PS1.3.020 "Sécurité et Qualité des Produits Aquacoles le Développement d'une Voie Commune Tuniso-Sicilienne" co-financed by The European Union. We would like to thank Dr. Slim TRITAR from FMB (Ferme marine de Bizerte) for providing the samples.

### BIBLIOGRAPHY

- AOAC (1990) (Association of official analytical chemists). Official methods of analysis. 15th Ed. Hellrich K. (ed.) Agricultural chemicals; contaminants; Drugs.,1.
- AUBOURG S.P., PINEIRO C., GALLARDO J.M & BARROS-VELAZQUEZ J. (2005). Biochemical changes and quality loss during chilled storage of farmed trout (*Psetta maxima*). Food Chem., 90: 445-452.
- AVEIRO M., PELLIZZARO Q.C., AMBONI R.D.M.C., BATISTA C.R.V., BEIRÃO H.L & BARRETO. P. L. M. (2007): Chemical, microbiological and sensory changes of marinade mussel (*Perna perna* linné 1758) storage at 4°C. Alim. Nutr., Araraquara.,18:121-126.
- BALLESTEROS S. A., CHIRIFE, J & BOZZINI J. P. (1993). Specific solute effects on Staphylococcus aureus cells subjected to reduced water activity. *Int. J. Food Microbiol.*, 20:51-66.
- BAO D.N.H., ARASON S & PORARINSDOTTIR K.A. (2007). Effects of dry ice and superchilling on quality and shelf life of arctic charr (*Salvelinus alpinus*) fillets. *International Journal of Food Engineering*, 3:1-27.
- BEUCHAT L. R. (1983). Influence of water activity on growth, metabolic activities and survival of yeasts and molds. *J.Food Protect.*, 46:135-141.
- BILGIN S., ERDEM M.E & DUYAR H.A. (2006). Chemical quality changes of brown shrimp *Crangon crangon* (Linnaeus, 1758), stored at refrigerated temperatures as boiling and raw. *Science and Engineering Journal of Firat University.*, 18:171-179.

- BRANNAN R. G & ERICKSON M. (1996). Sensory assessment of frozen stored channel catfish in relation to lipid oxidation. *Aquatic Food Technology.*, 5: 67-80.
- CADUN A., CAKLI S & KISLA D. (2005). A study of marination of deepwater pink shrimp (*Parapenaeus longirostris*, Lucas, 1846) and its shelf life. *Food Chemistry.*, 90 (1-2): 53-59.
- CADUN A., KISLA D & CAKLI S. (2008). Marination of deep-water pink shrimp with rosemary extract and the determination of its shelf life. *Food chem.*, 109: 81-87
- CAKLI S., KLINIC B., DINCER T & TOLASA S. (2006). Effect of using slurry ice during transportation on the microbiological, chemical and sensory assessment of aquacultured sea bass (*Dicentrarchus labrax*) stored at 4°C. Critical *Review in Food Science and Nutrition.*, 46: 453-458.
- CABRER A.I., CASALES M., YEANNES M.I. (2002). Physical and chemical changes in anchovy (*Engraulis anchoita*) flesh during marination. Journal of Aquatic Food Product Technology., 11(1), 19-31.
- CEC. (1995). Commission of the European Comity. Decision 95/149/EC of 8 march 1995 fixing the total volatile basic nitrogen (TVB-N) limit values for certain categories of fishery products and specifying the analysis methods to be used Brussels
- DALGIÇ G. & ERKOYUNCU I. (2003). The quality changes in the smoked mussels (*Mytilus* galloprovincialis) marinades. Süleyman Demirel Üniv. Egirdir Su Ürûnleri Fak. Dergisi., 2(10): 20-25
- DGSV.(2011): mise à jour du 05/12/2011 de l'annexe 1 de la NS. n° 200/2039 du 21/08/2006, Direction Générale des Services Vétérinaires, Ministère de l'Agriculture, Tunisie.
- DGPA. (2015): Direction Générale des Services Vétérinaires, Ministère de l'Agriculture, Tunisie.
- DUYAR H.A., GARGACI A & ALTINELATAMAN C. (2012). Determination of chemical composition and shelf life of shad (*Alosa tanaica* Grimm.1901) in refrigeration condition. *Journal of Fisheries Science*.com., 6:1-8.
- ERKAN N. (2005). Changes in quality characteristics during cold storage of shucked mussels (*Mytilus galloprovincialis*) and selected chemical decomposition indicators. J. Sci.Agric., 85:2625-2630.
- FEENEY R.E. (1977). Chemical changes in food proteins In: Evaluation of Proteins for Humeans. Bodwwell, C.E (Ed.). Connecticut: The AVI Publishing Company, Inc., 233-251.

- FOLCH J., LEES M., STANLEY G.H.S. (1957). A simple method for the isolation and purification of total lipids form animal tissue. *J. Biol. Chem.*, 226: 497-509.
- GENOT C. (1996). Some factors influencing TBA test. In Report of diet-ox project (AIRIII-CT-92-1577).
- GIRARD J.P & Valin C.(1988). Technologie de la viande et des produits carnés. Technique et documentation Lavoisier, Paris.117-135.
- GOULAS A.E & KONTOMINAS M.G. (2005). Effect of salting and smoking method on the keeping quality of chub mackerel (*Scomber japnicus*): Biochemical and sensory attributes. *Food Chem.*, 93: 511-520.
- GOULD G. W. (1985). Osmoregulation: is the cell just a simple osmometer ? The microbiological experience, in A Discussion Conference: Water Activity: A Credible Measure of Technological Performance and Physiological Viability? Faraday Div., R. Soc. Chem., Girton College, Cambridge, England, July 1-3.
- GOULD G. W & Christian J. H. B. (1988).
  Characterization of the state of water in foods
  biological aspects, in Food Preservation by Moisture Control, C. C. Seow, T. T. Teng, and C. H. Quah, Eds., *Elsevier*, New York. 43-56.
- GOPAL T.K.S., BALACHANDRAN K.K., SURENDRAN P.K & GOVINDAN T.K.(1985) development of flexible packaging for mussel pickled in oil. *In Harvest and Post Harvest Technology of fish.*,744.
- GULDAS M., HECER C. (2010). Influences of the selected additives on the weight loss and organoleptic properties of marinated mussels and squids. ACTA VET. BRNO 81: 263-267.
- HARTREE E. F. (1972). Determination of protein: a modification of the Lowry method that gives a linear photometric response. *Analytical biochemistry*, 48(2), 422-427.
- HARRIGAN W.F & McCANCE M.E. (1976) Laboratory Methods in Microbiology, Academic press, London and New York.
- HUSS H. H. (1988). Fresh fish quality and quality changes. Rome: Food and Agriculture Organization of the United Nations.
- HUSS H. H. (1995). Quality and quality changes in Fresh Fish.In: *FAO Fisheries Technical Paper*, N°348., Rome. Italy, FAO
- HWANG C & TAMPLIN M.L. (2005). The influence of mayonnaise pH and storage temperature on the growth of Listeria monocytogenes in seafood salad. *International Journal of Food Microbiology.*, 102: 277-285.
- ICMSF (1986) (International Commission on Microbiological Specifications for Foods), Microorganisms in Foods. Sampling for Microbiological Analysis: Principles and

Scientific Applications, Vol. 2, 2nd ed. University of Toronto Press, Toronto, pp. 181– 196.

- ICMSF.(1993) International Commission on Microbiological Specification of Foods. Microorganisms in foods 2. Sampling for microbiological analysis. Principles and applications, 2<sup>nd</sup> edn. Toronto, Canada: University of Toronto Press
- JARVIS M. J., TUNSTALL-PEDOE H., FEUERABEN C., VESEY C & SALOOJEE Y. (1987). Comparison of tests used to distinguish smokers from non-smokers. *American Journal of Public Health.*, 77(11)/ 1435-1438.
- KAYA G. K. & BASTURK Ö. (2015). Determination of some quality properties of marinated sea bream (*Sparus Aurata* L., 1758) during cold storage. *Food Science and Technology* (Campinas), 35(2), 347-353.
- KILINC B & CAKLI S. (2004). Chemical microbiological and sensory changes in thawed frozen fillets of sardine (*Sardina pilchardus*) during marination. *Food Chem.*, 88:275-280.
- KILINC B & CAKLI S. (2005). Determination of the shelf life of sardine (*Sardina pilchardus*) marinades in tomato sauce stored at 4°C. *Food control.*, 16:639-644.
- KOUTSOUMANIS K & NYCHAS G.J.E. (1999). Chemical and sensory changes associated with microbial flora of Mediterranean boque (*Boops boops*) stored aerobically at 0, 3, 7 and 10°C. *Applied and Environ. Microbiol.*, 65: 698-706.
- KYRIAZI-PAPADOPOULOU A., VARELTZIS K., BLOUKAS J.G & GEORGAKIS S. (2003). Effect of smoking on qulity characteristics and shelf-life of Mediterranean mussel (*Mytilus* galloprovincialis) meat under vacuum in chilled storage. *Ital. J. Food Sci.*, 3(15): 371-381.
- LEISTNER L., Rodel, W & Krispien, K. (1981). Microbiology of meat and meat products in high and intermediate moisture ranges. L.B. Rockland, G.F. Stewart (Eds.), Water Activity: Influences on Food Quality, Academic Press, New York. 855-91.
- MAKTABI S., ZAREI M & CHDORBAF M. (2015). Effect of Traditional marinating on bacterial and chemical characteristics in frozen rainbow trout fillet. *Journal of food quality and hazards control.*, 2: 128-133.
- McLAY B.R. (1972). Marinades. Ministry of Agriculture Fisheries and Food. Tony Advisory Note, 56: 14.

- MURRAY J., & BURT J. R. (1969). The Composition of Fish. Torry Advisory Note No. 38, Aberdeen, UK: Torry Research Station.
- ORBAN E., DI LENA T., CASINI I., MARZETTI A & CAPRONI R. (2002). Seasonal changes in meat content, condition index and chemical composition of mussel (*Mytilus* galloprovincialis) cultured in two different Italian sites. Food chem., 77: 57-65.
- OZDEN Ô. (2005). Change in amino acid and fatty acid composition during shelf-life of marinated fish. *Journal of the Science of Food and agriculture.*, 85(12): 20015-20020.
- OZGUL Y., OZGUL F & KULEY E. (2010). Effects of combining of smoking and marinating on the shelf life of anchovey stored at 4°C. *Food science and Biotechnology.*, 19:69-75.
- OZGUL Y & BALIKCI E. (2013). Effect of Various processing methods on quality of Mackerel (*Scomber scomber*). *Food Bioprocess Technol.*, 6: 1091-1098.
- OZYURT G., KULEY E., BALIKCI E., KACAR C., GOKDOGAN S., ETYEMEZ M & OZGUL F. (2012). Effect of the icing with rosemary extract on the oxidative stability and biogenic amine formation in sardine (*Sardinella aurita*) during chilled storege. *Asian Journal of Food* and Bioprocess Technology., 5: 2777-2786.
- PAKAWATCHAI C., SIRIPONGVUTIKORN S & USWAKESMANEE W. (2009). Effect of herb and spice paste on the quality changes in minced salmon flesh waste during chilled storage. *Asian Journal of Food and Agro-Industry.*, 2: 481-492.
- PEZESHK S., REZAEI M., RASHEDI H & HOSSAINI H. (2012). Investigation of antibacterial and antioxidant activity of turmeric extract (*Curcuma longa*) on rainbow trout (*Oncorhynchus mykiss*) in vitro.Iranian *Journal of Food Science and Technology.*, 35:77-87.
- PIRINI M., GATTA P.P., TESTI S., TRIGARI G & MONETTI P.G. (2000). Effect of refrigerated storage on muscle lipid quality of sea bass (*Dicentrarchus labrax*) fed on diets containing different levels of vitamin E. *Food Chemistry.*, 68: 289-293.
- RAHMAN M S & LABUZA T P. (1999). Water activity and food preservation. In: Handbook of food preservation (ed. M.S. Rahman). Marcel Dekker, New York, 173-432.
- REGOST C., JAKOBSEN J.V & RORA A.M.B. (2004). Flesh quality of raw and smoked fillets of Atlantic salmon as influenced by dietary oil sources and frozen storage. *Food Research International.*, 37: 259-271.

- RUIZ-CAPILLAS C & HORNAR W.F.A. (1999). Determination of the Trimethylamine and total volatile basic nitrogen in flesh fish by flow Injection analysis. *Journal of the Science of Food and Agriculture.*, 14(79): 1982-1986.
- RUIZ-CAPILLAS C & MORAL A. (2001). Correlation between biochemical and sensory quality indices in hake stored in ice (2001a).*Food Research International.*,34:441-447
- RUIZ-CAPILLAS C & MORAL A. (2005). Sensory and biochemical aspects of quality of whole big eye tuna (*thunnus obesus*) during bulk storage in controlled atmospheres. Food in controlled atmospheres. Food chemistry, 89,347-354.
- SALLAM K.I., AHMED A.M., ELGAZZAR M.M & ELDALY E.A. (2007). Chemical quality and sensory attributes of marinated Pacific saury (*Cololabis saira*) during vacuum- packaged storage at 4°C. *Food Chem.* 102: 1061-1070.
- SAINCLIVIER M. (1985). Des techniques ancestrales à leurs réalisations contemporaines: salage, séchage, fumage, marinage, hydrolysats. In: L'industrie alimentaire halieutique. II., Rennes: Vol. Bulletin scientifique et technique de l'école nationale supérieure agronomique et du centre de recherches de Rennes, 1985, 6-162.
- ŞENGÖR G.F., GUN H & KALAFATOGLU H. (2004). The determination of microbial flora water activity and chemical analyses in smoked, canned mussel (*Mytilus* galloprovincialis). Turk.J. Vet.Anim. Sci. 28:793-797.
- SILVERMAN G.J., MUNSEY D. T., LEE C & Ebert E. (1983). Interrelationship between water activity, temperature and 5.5 percent oxygen on growth and enterotoxin A secretion by Staphylococcus aureus in precooked bacon. J. Food Sci., 48:1783-1786, 1795.
- SMALL A.C & VAN STRALEN. (1990). Average annual growth and condition of mussels as function of food source. *Hydrobiologia.*, 195: 179-188.
- STAMATIS N & VAFIDIS D. (2009). Effect of marinating and vacuum storage at 6°C on the fate of chemical, microbial and sensory quality indices of echinoid gonads *Paracentrotus lividus* Lamark, 1816. *International Jouranl of Food Science and Technology.*, 44(8): 1626-1633.
- TARLADGIS B., WATTS B.M & YONATHAN M. (1960). Distillation method for determination of malonaldehyde in rancid food. *Journal of American Oil Chemistry Society.*, 37: 44-48.
- TURAN H., SOMMEZ G., CELIK Y.M., YALCIN M & KAYA Y. (2006). Effect of different

salting process on the storage quality of Mediterranean mussel (*Mytilus* galloprovincialis L.1819). Journal of Muscle Foods., 18:380-390.

- TURAN H., SÖNMEZ G., CELIK Y.M., YALCIN & KAYA Y.(2008) The effect of hot smoking on the chemical composition and shelf life of Mediterranean mussel (Mytilus galloprovincialis) under chilled storage. *Journal of food processing and preservation.*, 32(6), 912-922.
- VARELTZIS K., ZETOU F & TSIARAS I. (1993). Textural deterioration of chub mackerel (*Scomber japonicas collias*) and smooth jound (*Mustelus mustelus* L.) in frozen storage in

relation to chemical parameter. *Lebensmittel-Wissenshaft Und-Technologie*, 21:206-2011.

- VARLIK C., UğGUR, M., GOKOGLU N & GUN, H. (1993). Principles and methods of quality control in seafood. İstanbul: *Food Technology Association.*,174.
- YASHODA K.P., RAO R.J., MAHENDRAKAR N.S & RAO D.N. (2005). Marination of sheep muscles under pressure and its effect on meet texture quality. *Journal of Muscle Foods.*, 16: 184-191.
- YEANNES M.I & CASALES M.R. (2008). Modification in the chemical compounds and sensorial attributes of Engraulis anchoita fillet during bringing and marinating stages. *Ciência e Technologia de Alimentos.*, 28(4): 798-803.